

Protocol: Agarose Pads for Microscopy

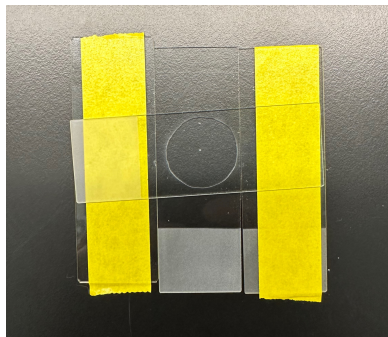
It is useful in many applications to immobilize specimens suspended in liquid on a microscope slide, such as yeast and *C. elegans*. Follow this protocol to make agarose pads for microscopy.

What is needed:

Agarose

Glass Pasteur pipette

1. Make a solution of 2% agarose in H₂O (for example, add 1g of agarose to 50ml of water).
2. Microwave until the solution goes completely clear. Check the solution periodically and swirl. Careful- agarose solutions can boil over when swirled. Be gentle and use caution.
3. Apply lab tape to two microscope slides (or reuse microscope slides for this purpose). Place a clean microscope slide between the two taped slides.
4. Use a glass Pasteur pipette and rubber bulb to apply 4 drops of hot 2% agarose solution to the center of the clean slide. Gently drop a second clean slide on top, making the cross pattern below. This sandwiches the agarose solution into a thin layer with the thickness of the lab tape.



5. Allow the agarose to cool for about 3 minutes. Then gently pull, twist, or slide the two clean microscopes apart such that the agarose pad adheres to one of the two clean slides.
6. Mount specimens by applying them to the agarose pad. Add a drop of VectaShield mounting medium to the specimen (if desired). Apply a coverslip and seal with nail polish.